USE OF LOW-OXYGEN ATMOSPHERES IN QUARANTINE APPLICATIONS TO ERADICATE MUS MUSCULUS, THE HOUSE MOUSE

JULIE A. CASSELLS¹, J.E. VAN S. GRAVER¹, R.H. BURTON² AND H.J. BANKS¹

¹Stored Grain Research Laboratory, CSIRO Division of Entomology,

GPO Box 1700, Canberra ACT 2601, Australia

²Cooperative Bulk Handling, GPO Box L886,

Perth WA 6001, Australia

ABSTRACT

The feasibility of using low-oxygen (O_2) atmospheres to eradicate infestations of mice (*Mus musculus*) in order to meet quarantine standards was investigated. Drilling equipment and transportable buildings were enclosed within standard plastic fumigation sheets both with and without floor sheets. The enclosures, 45–189 m³, were dosed with a low- O_2 atmosphere ($\approx 1.5\%$ O_2) generated by a membrane nitrogen separation system. Purging was observed to follow a simple free mixing model in form. The atmospheres were created using about 20% less gas than the amount predicted on the basis of free mixing alone. Applying this model allows selection of generators of suitable size. Leakage from sheeted containers was very slow after purging (0.04% d⁻¹ or less), showing that a high degree of gas retention was easily achieved.

Atmospheres containing less than 2.5% O₂ were obtained and maintained within the enclosures for a minimum of 6 h. Published literature suggests this would have caused 100% mortality of exposed mice. Work is continuing to establish the most cost-effective exposure regime.

Application of this treatment as a quarantine measure against rodents is discussed and its costs compared with conventional methods of disinfestation using methyl bromide.

INTRODUCTION

In 1994, the Stored Grain Research Laboratory was commissioned by West Australian Petroleum Pty Limited (WAPET) to find an effective replacement for methyl bromide (MB) for control of rodents in its quarantine operations. This chemical has been used for rodent control world-wide since the 1940's (Borg, 1944; Lembrez, 1966) and is still internationally recommended for this purpose (Bond, 1984).

WAPET is an oil and gas company for exploration and production, operating on two islands on the northwest shelf of Australia. Both islands, Barrow and Thevenard, are nature reserves serving as refuges for rare or endangered wildlife. Consequently, the

Company's operations are constrained by both voluntary controls and those mandated by the Environmental Protection Authority on imported material, including plants and animals.

The company, which has won industry recognition for its environmentally friendly operations, has gained two awards for its environmental policies. Even after 30 years of operation, Barrow Island is still a refuge for a number of animal species, such as the Burrowing Bettong, that have disappeared from their original habitats on the mainland. If rabbits, cats, foxes, rats, mice, snails or weeds were allowed to enter these islands, they could seriously, perhaps permanently, disturb the ecology of the native wildlife, possibly causing extinctions through predation and deprivation.

WAPET enforces a quarantine program to ensure that exotic animals, specifically mice, are not carried from the mainland to the islands. Equipment that can be infested by rodents and other vermin is subjected to MB fumigation. Items such as transportable offices, accommodation units, tool sheds, equipment used for food storage, mobile messes, etc. are commonly treated before they are shipped from the mainland to the islands. Also, because non-native mice have become established on Thevenard Island, similar restrictions apply when equipment is transported from there to Barrow Island.

In keeping with WAPET's environmental concerns, and with occupational health and safety concerns, the company desired to replace MB for this particular application. However, the company wanted any potential replacement for MB to be environmentally safe; safer for workers than the existing system; compatible with exploration and production operations for oil and gas; compatible, in terms of speed of operation, with the existing method of disinfestation; and suitable for use at remote locations.

After visiting and inspecting WAPET's on- and off-shore operations it was concluded that three potential alternatives could be considered. These were use of heat/high temperatures, high carbon dioxide (CO₂) atmospheres and low-oxygen (O₂) atmospheres.

Heat as an alternative

This was ruled out because of potential damage to the heat sensitive equipment used during the company's operations. More importantly, it posed a potential, and serious, fire hazard when used in the vicinity of gas and oil, whether wells or storage vessels.

Carbon dioxide as an alternative

Early reports of the potential use of CO_2 in the storage environment to control mice and other rodents were published by Pieniazek and Christopher (1947) and Southern (1954). It has been used to control mice both in seed and feed stores and in cold storages, for which an atmospheric concentration of 25% CO_2 was applied for periods extending up to 2 h (Hamel, 1986). It is widely used in euthanasia for killing a wide range of animals, including rats and mice (Andrews *et al.*, 1993; Hansen *et al.*, 1991; Hornett and Haynes, 1984). However, its use in this particular application was constrained by the logistics and costs involved in transporting the large quantities of CO_2 that would be required.

Low-O₂ atmospheres as an alternative

Use of nitrogen (N_2) to generate low- O_2 atmospheres was considered most appropriate because the proposed system met most of WAPET's replacement criteria. N_2 is already used in oil and gas exploration and production to remove air in tanks and pipes in order to prevent fires and explosions. A membrane generation system was chosen because such systems are reliable, small and portable, and they are already used by the industry elsewhere. Also, in our experience, low- O_2 atmospheres are more easily contained in plastic enclosures sealed with sand or sand snakes than are high CO_2 atmospheres.

 N_2 is widely used for euthanasia of a wide range of animals, including rats and mice (Andrews *et al.*, 1993). Rats collapse in approximately 3 min, and stop breathing in 5–6 min, in low- O_2 atmospheres produced with N_2 flowing at a rate of 39% of chamber volume per minute, thus falling to about 3% O_2 after 5 min (Hornett and Haynes, 1984). For mink, in 100% N_2 the time to death is 134 sec, but Hansen *et al.* (1991) recommend an extended exposure period of 5 min. Dogs are killed in less than 5 min by N_2 atmospheres containing 1.5% O_2 (Herin, 1978; Quine, 1980). When a group of 72 animals consisting of cats, kittens, rabbits and dogs was placed in a N_2 atmosphere (final O_2 concentration declining to $\leq 4.0\%$), 70 collapsed within 1 min of the O_2 concentration falling to 10%; 66 suffered respiratory arrest within 2 min of collapsing; and 66 had no detectable heart beat at 6 min, or less, after the O_2 concentration fell to <10% (Quine *et al.*, 1988).

However tolerance to hypoxia, in the short exposure periods used for euthanasia, has been observed in newborn mice, rats, rabbits, kittens and puppies (Blackmore, 1993; Herin, 1978; Quine, 1980; Quine *et al.*, 1988).

Barrere (1980) reported that atmospheres containing 99% N_2 killed rodents in less than half an hour in grain storages, but he gave no indication of a specific dosage regime for these pests. Pryor *et al.* (1974) in working with mice found that deaths occurred at 7.5% O_2 in 4 h experiments. Levin *et al.* (1987) calculated LC₅₀ values for rats exposed to low- O_2 atmospheres for 30 min in an atmosphere of 7.5% O_2 .

This paper reports on a trial undertaken to assess the feasibility and effectiveness of using low- O_2 atmospheres to eradicate infestations of mice in equipment used by the oil and gas exploration and production industry.

MATERIALS AND METHODS

The equipment disinfected during this trial consisted of drilling pipes (in racks), a toolshed container, an equipment container and transportable offices both with and without wheels. Dimensions and volumes of the seven lots of equipment after enclosure under fumigation sheets are given in Table 1. The sheets used were made from either yellow nylon reinforced laminated PVC or black Valeron (cross laminated HDPE). All equipment was positioned on cement stabilised sand prior to being enclosed and disinfested.

To obtain an indication of gas loss downward through the ground, three of the seven treatments were carried out using a gastight floor sheet. This was laid on the ground and the equipment placed on it before being enclosed and sealed for treatment.

TABLE 1
Dimensions and volumes of equipment treated, and type of enclosure sheet used

Treat		Dimensions (m)					
ment no.	Equipment	length	width	height	Volume (m ³)	Fumigation sheet	Ground sheet
1	Pipe racks 1	10.2	5	1.1	56	Valeron	
2	Pipe racks 2	10.2	5	1.1	56	Reinforced laminated PVC	_
3	Container 1; Toolshed	6.0	3	2.5	45	Valeron	-
4	Container 2; Kumo shack	12.2	3	2.5	92	Valeron	-
5	Container 3; Transportable office on wheels	13.5	4	3.5	189	Valeron	Valeron
6	Container 4; Jan's office rooms	13.5	3	3.1	126	Reinforced laminated PVC	Valeron
7	Container 5; Transportable office on wheels	13.5	4	3.5	189	Valeron	Valeron

Four pipe racks were treated. These were enclosed as pairs under a fumigation sheet, and each pair was disinfested as a single unit. All other items were enclosed and treated as single units. Sharp edges and protrusions from the equipment were covered to prevent their penetrating and tearing the fumigation sheets. The fumigation sheets were sealed and anchored to the ground, or to the floor sheet, using sand heaped approximately 0.5 m high continuously along the perimeter of the sheeted enclosures at the base of the structures. All visible holes and tears in the fumigation sheets were patched or sealed prior to treatment. Ropes were tied around the middle of each enclosure to reduce billowing and consequent gas "pumping" in the wind.

 N_2 for the treatment was supplied from a Prism (Permea Inc., St Louis, Missouri, USA) N_2 membrane system supplied by Oxair Australia, Perth, WA. It had a rated output of $18 \text{ m}^3 \text{ h}^{-1}$ at $98\% N_2$ and consisted of an electrically powered compressor, an air storage tank, the membrane system and a N_2 receiving tank.

In this trial it was decided to obtain and hold the O_2 levels below 5.0% and maintain these levels long enough to initially ensure complete dispersal of the atmosphere throughout the equipment and thereafter to disinfest the equipment.

 N_2 for the treatments was piped from the generator via a 5-cm i.d. nylon-reinforced plastic pipe to a manifold fitted with three ball valves from which it was distributed to the enclosed equipment. These valves provided full (100%) flow with a 90° turn. For the purposes of this trial it was assumed that a 30° turn of the valve delivered 33.3% of the full flow and a 45° turn delivered 50% of the full flow. Subsequent calculations confirmed this assumption.

To prevent excess ballooning of the enclosures during the purge in treatments 1-3 (Table 1), gas was allowed to vent via a small (≈ 5 cm i.d.) hole in the sheeting approximately 1 m from the ground at the point furthest from the gas input line. In treatments 4 and 5 (Table 1), no vent was provided.

At the end of the treatment time, if the equipment was immediately required for shipment, the fumigation sheets were removed and the containers allowed to air until O_2 levels had been restored to 20-21%. Otherwise the containers remained enclosed until required. In such cases, the opportunity was used to monitor O_2 levels to determine the rate of leakage and permeation of air through the fumigation sheets after the N_2 purge was terminated.

O₂ concentrations achieved during the treatments were measured by sampling the atmosphere inside the enclosures. The samples were drawn out through 3-mm i.d. nylon tubing from various points in the enclosures (Figs. 1–4). The sampling lines were cleared

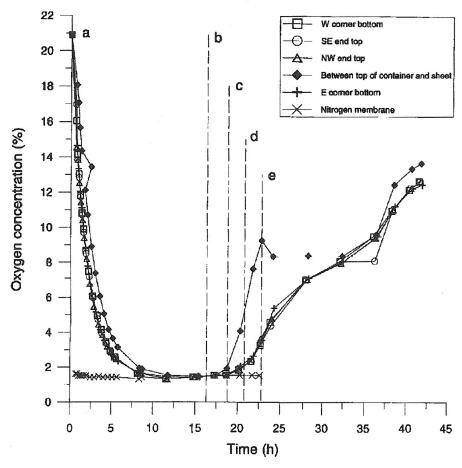


Fig. 1. O_2 concentrations during and after purge of container 1, the toolshed without floorsheet. Assumed flow rates during purge: a = 100%, b = 33%, c = flow rate reduced, d = flow rate reduced then increased and e = end of purge.

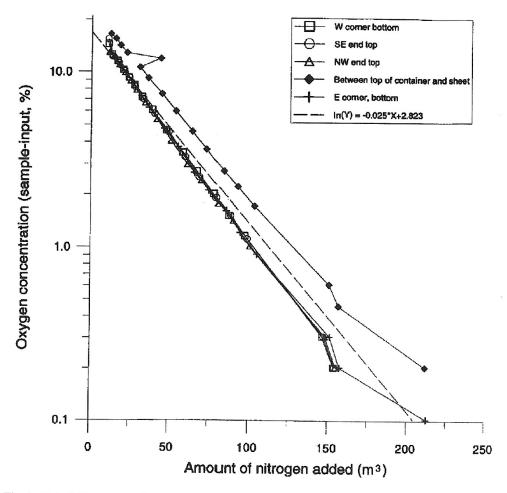


Fig. 2. Plot of O_2 concentration to show rate of O_2 decrease during purge of container 1, the toolshed without floorsheet.

by pumping for a period of 3–5 min at a rate of 150 ml min⁻¹ using a Masterflex (Barnant Co, Illinois, USA) peristaltic pump until a steady O_2 concentration reading was obtained. O_2 concentrations were measured using either an Otox 90 (Neotronics, Bishops Stortford, UK) or an Oxywarn 100 (Draeger) O_2 meter. During the treatments, the calibration of these meters was checked regularly against ambient air and industrial grade N_2 (BOC, $N_2 = 99.9\%$, $O_2 < 10$ ppm).

Ambient temperature as well as temperatures attained within the enclosures were measured using T-type copper/constantan thermocouples and recorded on a data logger (Data Electronics, Datataker DT100). Temperatures within the enclosures were measured at the same points from which gas samples were withdrawn (Figs. 1–4).

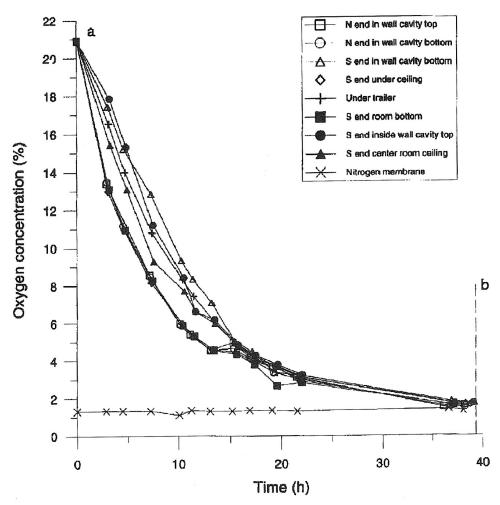


Fig. 3. O_2 concentrations during and after purge of container 5, transportable office on wheels with floorsheet. Flow rates during purge: a = 100% and b = end of purge.

RESULTS

The N_2 output from the membrane system provided an atmosphere with an O_2 concentration varying from 1.3–2.0% and averaging 1.5% O_2 (Figs. 1 and 3).

The O_2 concentrations achieved during the purges and the rate of O_2 decrease obtained are shown for container 1 (the toolshed without floor sheet) in Figs. 1–2 and for container 5 (the transportable office on wheels with floor sheet) in Figs. 3–4. The required O_2 concentrations ($\leq 5.0\%$) were achieved within a 5-h treatment period for container 1 and a 16-h treatment period for container 5, the volume of the latter being approximately 4 times that of the former (Figs. 1 and 3).

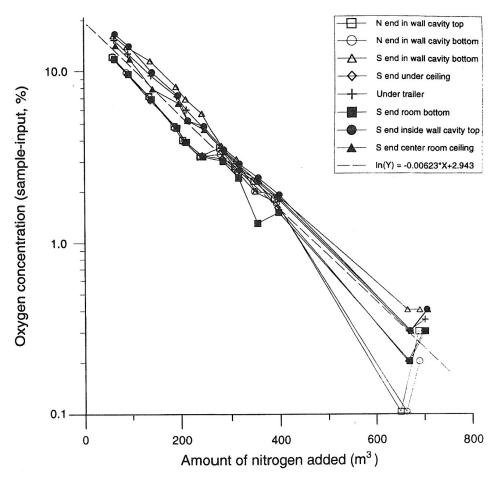


Fig. 4. Plot of O_2 concentration to show rate of O_2 decrease during purge of container 5, transportable office on wheels with floorsheet.

The amount of N_2 used during the purges was calculated from the rated output of the membrane system (18 m³ h⁻¹ at 98% N_2) and the degree of opening of the valves on the manifold regulating N_2 input into the enclosures. Table 2 compares the amount of N_2 required to reduce the O_2 level to 5% for each treatment as well as the rate and efficiency of the purge. Purging efficiency was calculated using Banks' (1979) formula, modified to take into account the fact that the purge gas, c_p , contained about 1.5% O_2 . This formula gives 100% efficiency if free mixing occurs throughout the space being purged:

Efficiency (%) =
$$100 \times \frac{volume\ of\ enclosure}{volumeof\ gas\ added} \times \ln \frac{21 - c_p}{5 - c_p}$$

The ventilation rate, k, a measure of the rate of gas loss from a system, was calculated

TABLE 2
Purge data for the seven treatments

Amount of

Treat ment no.	Equipment	Total N ₂ used (m ³)	Amount of N_2 required to reduce O_2 to 5% (m ³)	Treatment time at $O_2 \le 5\%$ (h)	Slope of pull- down line	Efficiency of purge ¹ (%)
1	Pipe racks 1	235	67	13	-0.0190	129
2	Pipe racks 2	165	55	6	-0.0187	157
3	Container 1; Toolshed	329	38	20	-0.0250	183
4	Container 2; Kumo shack	394	127	11	-0.0080	112
5	Container 3; Transportable office on wheels	416	211	12	-0.0048	138
6	Container 4; Jan's office rooms	406	116	27	-0.0086	168
7	Container 5; Transportable office on wheels	707	172	30	-0.0062	170

¹Calculated assuming 10% of the volume within enclosure filled with impervious objects.

using the formula given by Banks and Annis (1984),

$$\ln \frac{21 - c}{21 - c_0} = -kt$$

where c and c_0 are the O_2 concentrations at time t and at start of ventilation. Figure 5 shows the return of O_2 by natural leakage into the purge enclosure after the purge gas addition ceased.

The averages of the ambient temperatures and those attained within the enclosures during the treatments are given in Table 3.

DISCUSSION

The purge data for the seven treatments carried out during this trial are given in Table 2. In all cases, purging efficiencies in excess of 100%, as calculated on the basis of free mixing, were observed, suggesting there was some O_2 removal by direct displacement. Up to 50% less O_2 was needed than was expected.

Decrease in O_2 concentrations (Figs. 1, 3) during the purges was uniform at all points within the treated equipment. This included the wall and ceiling cavities of the transportable office. Even at roof height, areas which can provide nesting and breeding sites for rodents were subjected to O_2 concentrations $\leq 5\%$ during the exposure periods used here.

This trial demonstrated that it is feasible to reduce O_2 concentrations within sheeted enclosures efficiently to $\leq 5.0\%$ and to hold them at < 2.5% for periods extending to 6 h. In the work reported here, the objective was to attain low O_2 levels and maintain them

TABLE 3

Ambient temperature, and temperatures in sheeted equipment during treatment

Treat		Ambient temperature (°C)			Temperature inside enclosure (°C)		
ment no.	Equipment	Average	Average maximum	Average minimum	Average	Average maximum	Average minimum
1	Pipe racks 1	22.1	30.0	19.2	24.4	30.4	21.5
2	Pipe racks 2	22.5	30.1	19.2	23.8	29.6	21.2
3	Container 1; Toolshed	22.2	27.3	18.8	24.6	31.3	20.4
4	Container 2; Kumo shack	22.5	29.6	19.3	25.9	32.0	20.2
5	Container 3; Transportable office on wheels	21.9	25.9	20.1	25.2	29.2	18.4
6	Container 4; Jan's office rooms	20.0	27.3	17.4	20.8	29.3	18.7
7	Container 5; Transportable office on wheels	21.5	27.8	18.4	23.8	31.2	19.3

sufficiently long to kill any infestations of mice in the equipment treated. Levin *et al.* (1987) reported 100% mortality in 42 rats exposed for 30 min to atmospheres containing 7.5% O_2 , while Pryor *et al.* (1974, cited in Levin *et al.*, 1987) reported that mice were killed at the same O_2 concentration in 4 h experiments. Thus, with the O_2 concentrations achieved and the exposure periods used in this trial, it can be expected that at least all adult animals would have been killed. Similarly, we believe that the prolonged exposures to the low- O_2 atmospheres attained during this work, well in excess of those used for euthanasia, would have been lethal to neonate rodents.

With respect to a dosage and exposure regime, laboratory studies will be undertaken to confirm the results obtained during this trial and establish recommendations for atmospheric O_2 concentrations and exposure periods. In the absence of detailed mortality dosage data, no recommendation is made here. However, we expect that the required exposure periods will be substantially shorter than those in these trials.

It may be important for toxicological or logistical reasons to reduce the time needed for pull-down to low O_2 . This can be achieved by using a generator with a greater output than that used in this work. The modelling that was undertaken above allows for appropriate sizing of N_2 generators for a particular, specified pull-down.

Very low leakage as calculated by ventilation rates were observed in the three enclosures which were allowed to stand after purging. Two had a rate of about 0.04 per day, while the third admitted air at a rate of only only 0.008 per day (Fig. 5). The latter rate is very low, and this is perhaps attributable to the better seal afforded by the PVC and floor sheet.

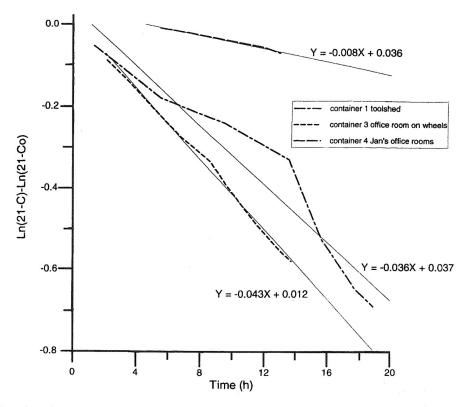


Fig. 5. Ventilation rate (O₂ decay) in enclosed equipment. Container 1 sheeted with Valeron without a floorsheet; container 3 with floorsheet; container 4 with PVC sheeting and a floorsheet.

Direct comparison of the cost of disinfestation with low-O₂ atmospheres to that of using MB is complex in this situation, where environmental and worker-safety concerns are difficult to quantify. However, the direct material costs involved in this use of low-O₂ atmospheres can be compared to those when MB is used. The cost of hiring the membrane system was AU\$ 500 as compared to the material cost of AU\$ 100 for MB, had it been used. Costs for electricity to run the membrane system are estimated at AU\$ 0.15 per m³ N₂. However, when MB is used, a licensed fumigator and protective equipment for the workforce engaged in this work are required. This would offset the cost difference, particularly in a remote region such as that in this trial series. Labour requirements for enclosing the equipment under gastight sheeting are the same in both cases.

MB treatments with a 4-h exposure period (Bond, 1984) followed by about 2-h airing time are typically rapid. Low- O_2 atmospheres are dependent on the capacity of the equipment available, which determines the rate of pull-down and the duration of the required exposure period. For the latter, we have not yet obtained definitive data, but we expect low- O_2 treatments to be at least as rapid as those with MB, given the appropriately sized generator and the mortality data reviewed above.

The time required for airing after treatment with low- O_2 atmospheres was shorter than for MB. The maximum time required with low- O_2 atmospheres was 0.5 h.

There are particular benefits to be gained by adopting the low-O₂ atmosphere technique in the application described here. Firstly, emissions of MB, a powerful ozone depletor (UNEP, 1992; WMO, 1995), would be reduced. Secondly, it would eliminate a worker-safety hazard from the oil and gas exploration and production workplace.

The technique may have wider application as a replacement for MB in such other situations as aircraft and other equipment already constructed to standards of gastightness. Furthermore, in situations where time constraints are critical, as with aircraft, there appears to be a case for adding CO₂ or carbon monoxide to increase the speed of action against rodents (Levin *et al.*, 1987; Pryor *et al.*, 1974).

CONCLUSION

The work described here demonstrated the feasibility of using low- O_2 atmospheres for rodent control in machinery and equipment which is under gastight fumigation sheeting. The technique using N_2 delivered from a membrane system to generate low- O_2 atmospheres is environmentally acceptable and safer than is the use of MB (Fig. 6). In operation, it is compatible with oil and gas exploration and production operations. It is also suitable for use at remote locations. The logistics of on-site gas generation provide environmental and

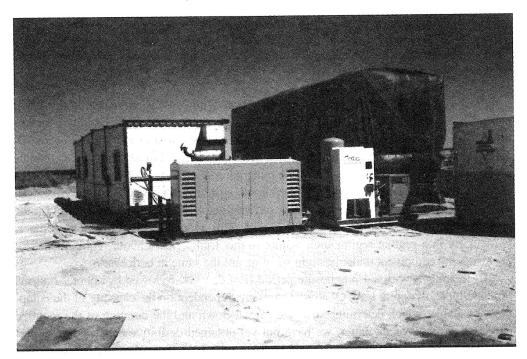


Fig. 6. View of membrane N2 system and enclosed transportable office on wheels under treatment.

worker-safety advantages that make it an attractive alternative to MB. The technique may have wider application beyond the oil industry.

ACKNOWLEDGEMENTS

We wish to thank our colleague, Geoff Russell, for drawing our attention to the literature on animal euthanasia. The following people all helped with or undertook literature searches on our behalf: Yvonne Hawkins (SGRL), Cdr T. Dickens and Capt Armando Rosales (Armed Forces Pest Management Board, Washington DC, USA), Lt Cdr Bob Gay (Defense Personnel Support Center Pacific, Alameda CA, USA) and Dr Dan Thompson (Denver Wildlife Research Center, USA).

At Thevenard Island, Rob Ward (CBH) assisted us. Andrew McCormack (WAPET) coordinated operations during the trial. We enjoyed, and are grateful for, the hospitality, enthusiastic interest and support that we received from WAPET staff and contractors.

We also thank Peter Annis and Chris Whittle for their helpful advice and criticism of the manuscript.

REFERENCES

- Andrews, E.J., Bennett, T.B., Clark, J.D., Houpt, K.A., Pascoe, P.J., Robinson, G.W. and Boyce, J.R. (1993) Report of the AVMA panel on euthanasia. *J. Am. Vet. Med. Assoc.* **202**, No. 2, 230–249.
- Banks, H.J. (1979) Recent advances in the use of modified atmospheres for store product pest control. In: Proc. 2nd Int. Working Conf. on Stored-Product Entomology, Ibadan, Nigeria, 1978, 198–217.
- Banks, H.J. and Annis, P.C. (1984) Importance of processes of natural ventilation to fumigation and controlled atmosphere storage. In: *Controlled Atmosphere and Fumigation in Grain Storages* (Edited by Ripp, B.E. *et al.*), Elsevier, Amsterdam, 299–323.
- Barrere, J.A. (1980) Use of nitrogen in cereal grain furnigation, storage. *Grain Age*, 9–10 July 1980, 12, 18.
- Blackmore, D.K. (1993) Euthanasia; not always Eu. Aust. Vet. J., 70 No. 11, 409-413.
- Bond, E.J. (1984) *Manual of Fumigation for Insect Control*. FAO Plant Production and Protection Paper No. 54, FAO, Rome, Italy. 432 pp.
- Borg, R.M. (1944) Methyl bromide as a fumigant for rats and mice in apple cold storages. *Proc. Amer. Soc. Hort. Sci.* **45**, 318–322.
- Hamel, D. (1986) Use of carbon dioxide for control of Mus musculus in storage. In: Proc. 4th Int. Working Conf. on Stored-Product Protection (Edited by Donahaye, E. and Navarro, S.), Tel Aviv, Israel, 21–26 September 1986, Maor-Wallach Press, Jerusalem, 636.
- Hansen, N.E., Creutzberg, A. and Simonsen, H.B. (1991) Euthanasia of mink (*Mustela vison*) by means of carbon dioxide (CO₂), carbon monoxide (CO) and nitrogen (N₂). *Br. Vet. J.* **147**, 140–146.
- Herin, R.A. (1978) Nitrogen inhalation as a method of euthanasia in dogs. *Am. J. Vet. Res.* **39**, No. 6, 989–991.
- Hornett, T.D. and Haynes, A.R. (1984) Comparison of carbon dioxide/air mixture and nitrogen/air mixture for the euthanasia of rodents: Design of a system for inhalation euthanasia. *Anim. Technol.* **35**, No. 2, 93–99.

- Lembrez, J. (1966) Rodent control on ships and in ports of Europe, Africa and the Middle East. *World Health Vector Control* **66**, No. 217, 123–128.
- Levin, B.C., Paabo, M., Gurman, J.L. and Harris, S.E. (1987) Effects of exposure to single or multiple combinations of the predominant toxic gases and low oxygen atmospheres produced in fires. *Fundam. Appl. Toxicol.* **9,** 236–250.
- Pieniazek, S.A. and Christopher, E.P. (1947) Carbon dioxide for controlling rodents. *Modern Refrigeration*, March 20, 69 and 84.
- Pryor, A.J., Fear, F.A. and Wheeler, R.J. (1974) Mass life fire hazard: Experimental study of the life hazard of combustion products in structural fires. *J. Fire Flammability: Combust. Toxicology Suppl.* 1, 191–235.
- Quine, J.P. (1980) Euthanasia by hypoxia using nitrogen: A review after four years of operation involving 20,500 animals. *Can. Vet. J.* 21, 320.
- Quine, J.P., Buckingham, W. and Strunin, L. (1988) Euthanasia of small animals with nitrogen; comparison with intravenous pentobarbital. *Can. Vet. J.* **29**, 724–726.
- Southern, H.N., ed. (1954) Control of Rats and Mice. Vol. 3: House Mice. Clarendon Press, Oxford, 199–215.
- UNEP (1992) Methyl Bromide: Its Atmospheric Science, Technology, and Economics. United Nations Environment Programme, Ozone Secretariat, Nairobi, Kenya. 41 pp.
- WMO (1995) Scientific Assessment of Ozone Depletion: 1994 (Executive Summary). World Meteorological Organization, Geneva, Switzerland, Global Ozone Research and Monitoring Project Report No. 37. 36 pp.